

# Molecular Residual Disease detection in early stage breast cancer with a personalized sequencing approach

536



The Royal Marsden  
NHS Foundation Trust

Rosalind Cutts<sup>1</sup>, Maria Coakley<sup>1</sup>, Isaac Garcia-Murillas<sup>1</sup>, Lara Ulrich<sup>2</sup>, Karen Howarth<sup>3</sup>, Warren Emmett<sup>3</sup>, Malcolm Perry<sup>3</sup>, Pete Ellis<sup>3</sup>, Charlene Knapé<sup>4</sup>, Stephen R. D. Johnston<sup>2</sup>, Alistair Ring<sup>2</sup>, Simon Russell<sup>5</sup>, Abigail Evans<sup>6</sup>, Anthony Skene<sup>7</sup>, Duncan Wheatley<sup>8</sup>, Mitch Dowsett<sup>9</sup>, Ian E. Smith<sup>2</sup>, Nicholas C. Turner<sup>1,2,9</sup>

1. The Breast Cancer Now Toby Robins Research Centre, The Institute of Cancer Research, London, UK, 2. Breast Unit, Royal Marsden Hospital, London, United Kingdom, 3. Inivata Ltd, Glenn Berge Building, Babraham Research Park, Cambridge, UK, 4. Inivata Inc, Research Triangle Park, NC, 5. Hinchingsbrooke Hospital, Hinchingsbrooke Park, Huntingdon, UK, 6. Poole General Hospital, Dorset, UK, 7. Royal Bournemouth Hospital, Bournemouth, UK, 8. Department of Oncology, Royal Cornwall Hospitals NHS Trust, Truro, UK, 9. The Ralph Lauren Centre for Breast Cancer Research, Royal Marsden Hospital, London, UK,

## Background

Detection of circulating tumor DNA (ctDNA) presents a strategy to identify Molecular Residual Disease (MRD) in patients with breast cancer. Tools capable of detecting ctDNA at lower concentrations are needed to increase sensitivity and lengthen lead time between ctDNA detection and relapse. We present results from a highly sensitive personalized sequencing approach for ctDNA detection of MRD based on multiple patient specific mutations.

## Methods

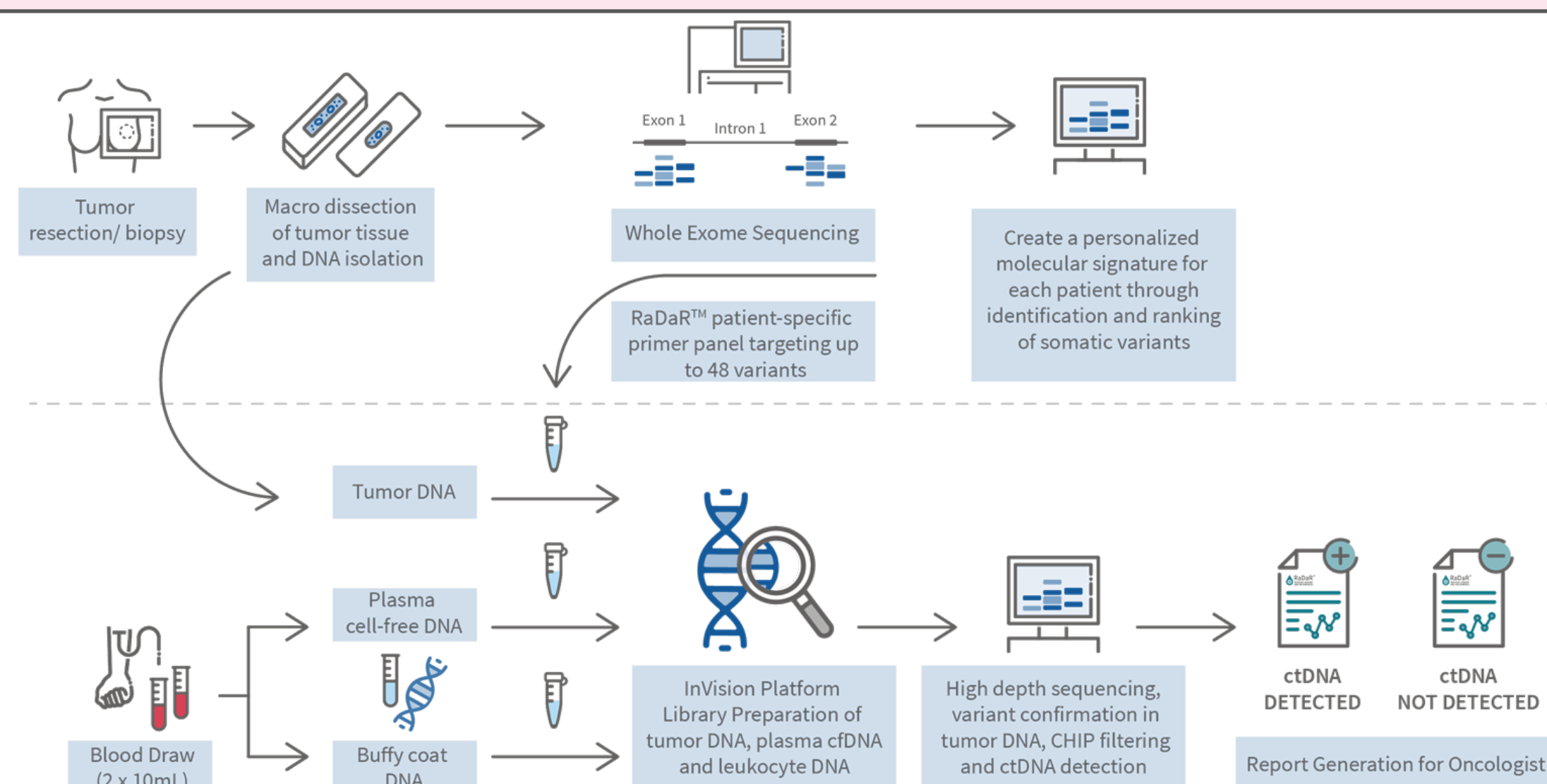
- 22 early breast cancer patients (12 HR+HER2-, 7 HER2+ and 3 TNBC) enrolled in the ChemoNEAR sample collection study were included.

- Tumor DNA from FFPE samples was Whole Exome Sequenced to identify patient specific mutations and design personalized Residual Disease and Recurrence (RaDaR™) multiplex PCR NGS assays.

- Cell free DNA was extracted from 147 plasma samples (median volume 4ml, range 0.5-5ml) and sequenced with RaDaR™ assays, with 10-61 variants (median 41) per panel, to 100,000x per locus. A matched single timepoint buffy coat was sequenced to verify removal of germline SNPs and any potential clonal hematopoiesis (CHIP). A proprietary algorithm was used to identify ctDNA. Droplet digital PCR (ddPCR) was performed as previously described<sup>1,2</sup>.

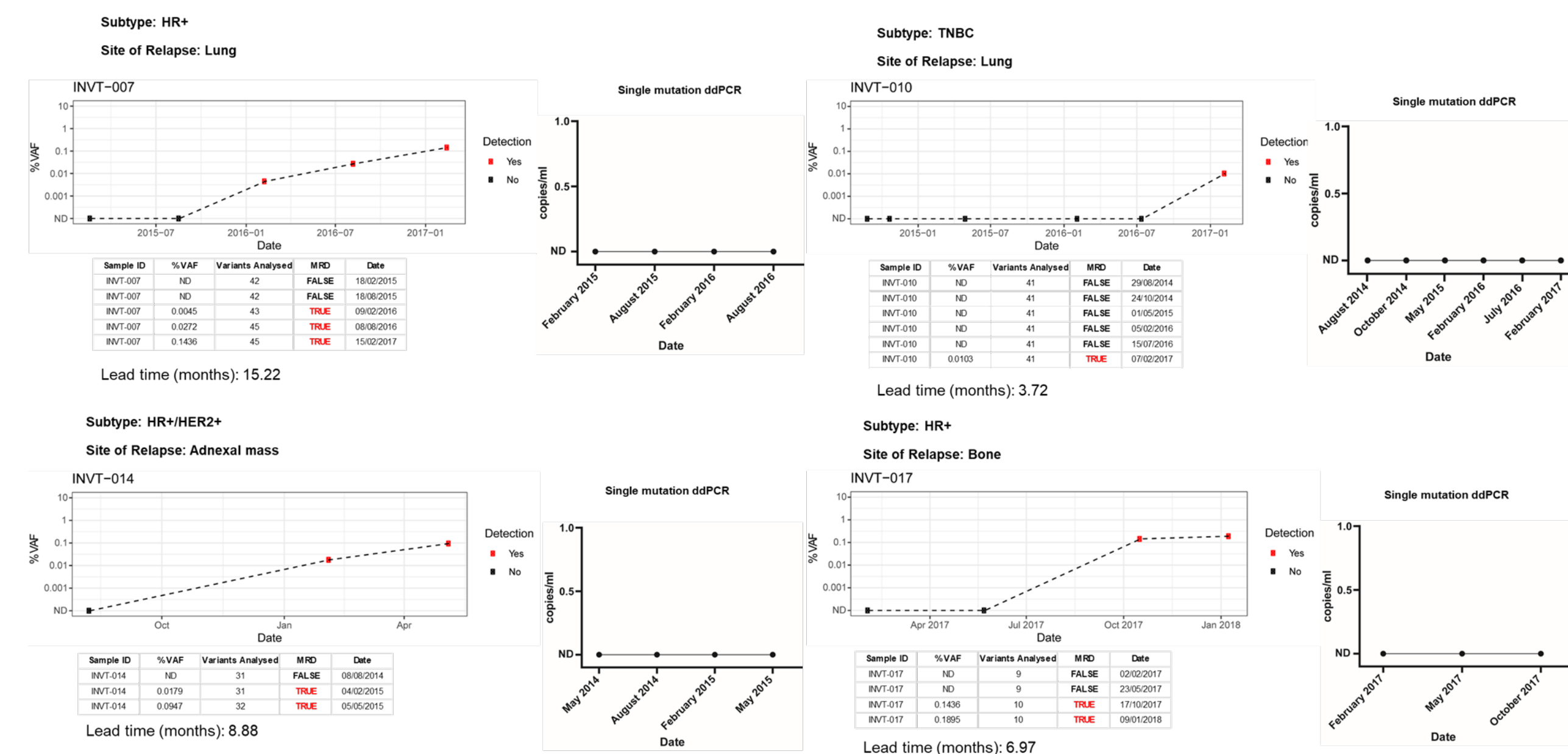
- Tumor Sequencing of multiple biopsy timepoints was carried out for 14 patients (mean 2.8 samples per patient) and clonal populations estimated with Pyclone. For clusters of greater than 10 mutations, RaDaR™ panels were supplemented with additional variants for clonal tracking.

## RaDaR™ assay workflow

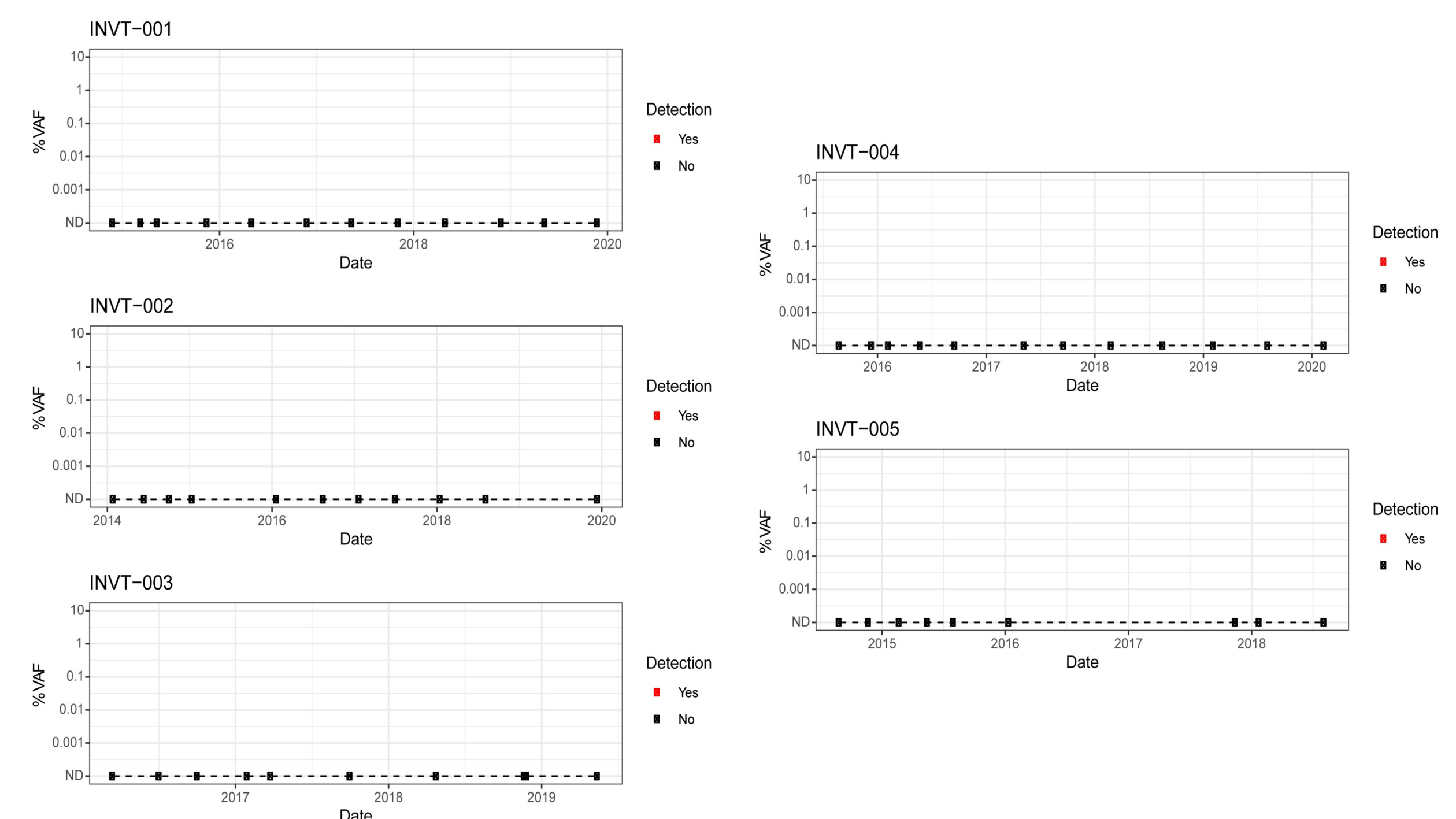


## MRD detection with personalized sequencing panels

MRD was identified in 100% (17/17) of relapsed patients (p=0.0002, Log rank test). Detection of ctDNA levels ranged from 0.0007% estimated Variant Allele Frequency (VAF) to 1.3% VAF (median 0.06% VAF). Five relapsed patients (4 extracranial and 1 brain only) had MRD detected with their corresponding RaDaR™ assays but not by single gene ddPCR.

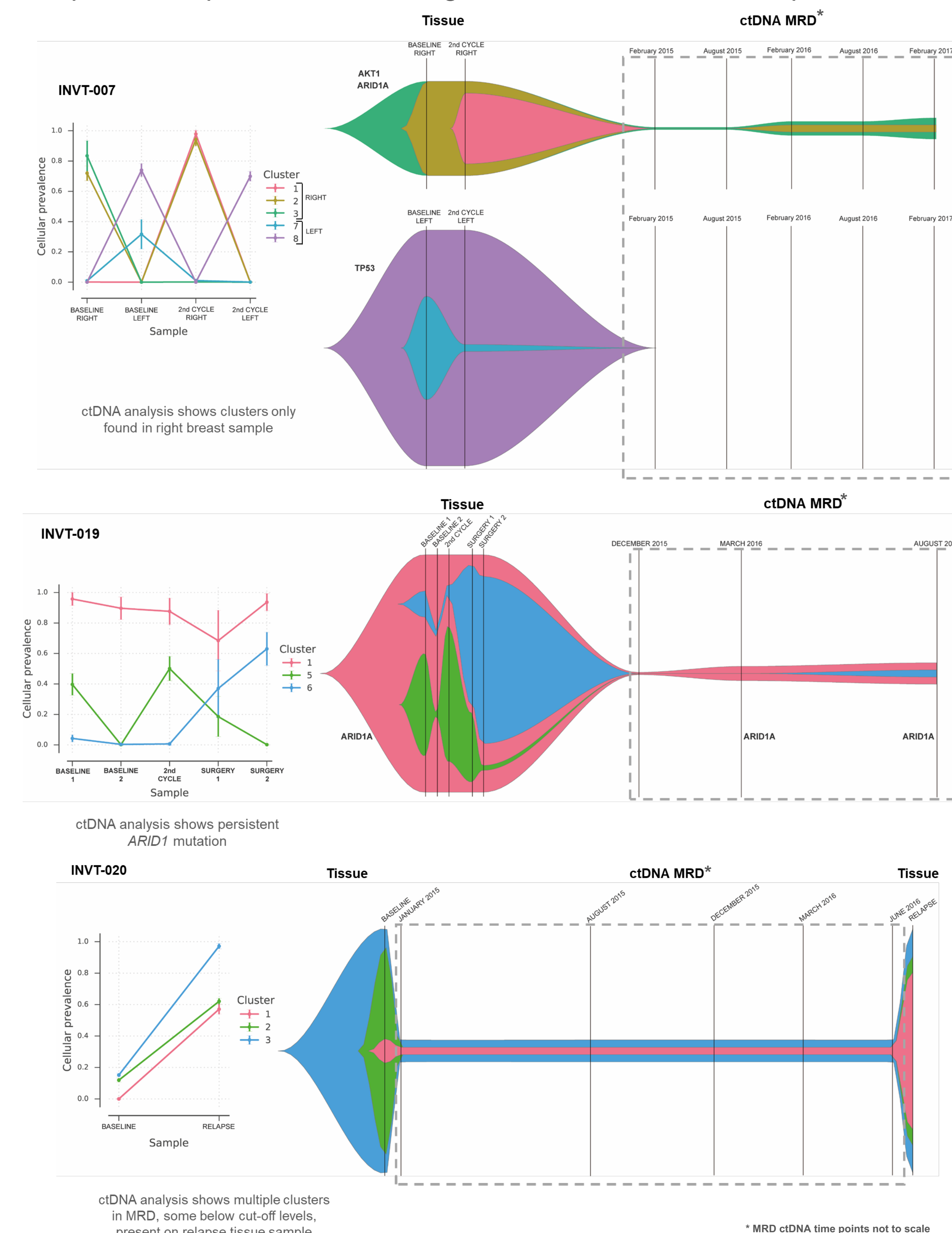


MRD was not identified in 54 time points (range 37.29 months to 69.22 months) in the 5 patients that did not relapse (p=0.0002, Log rank test).



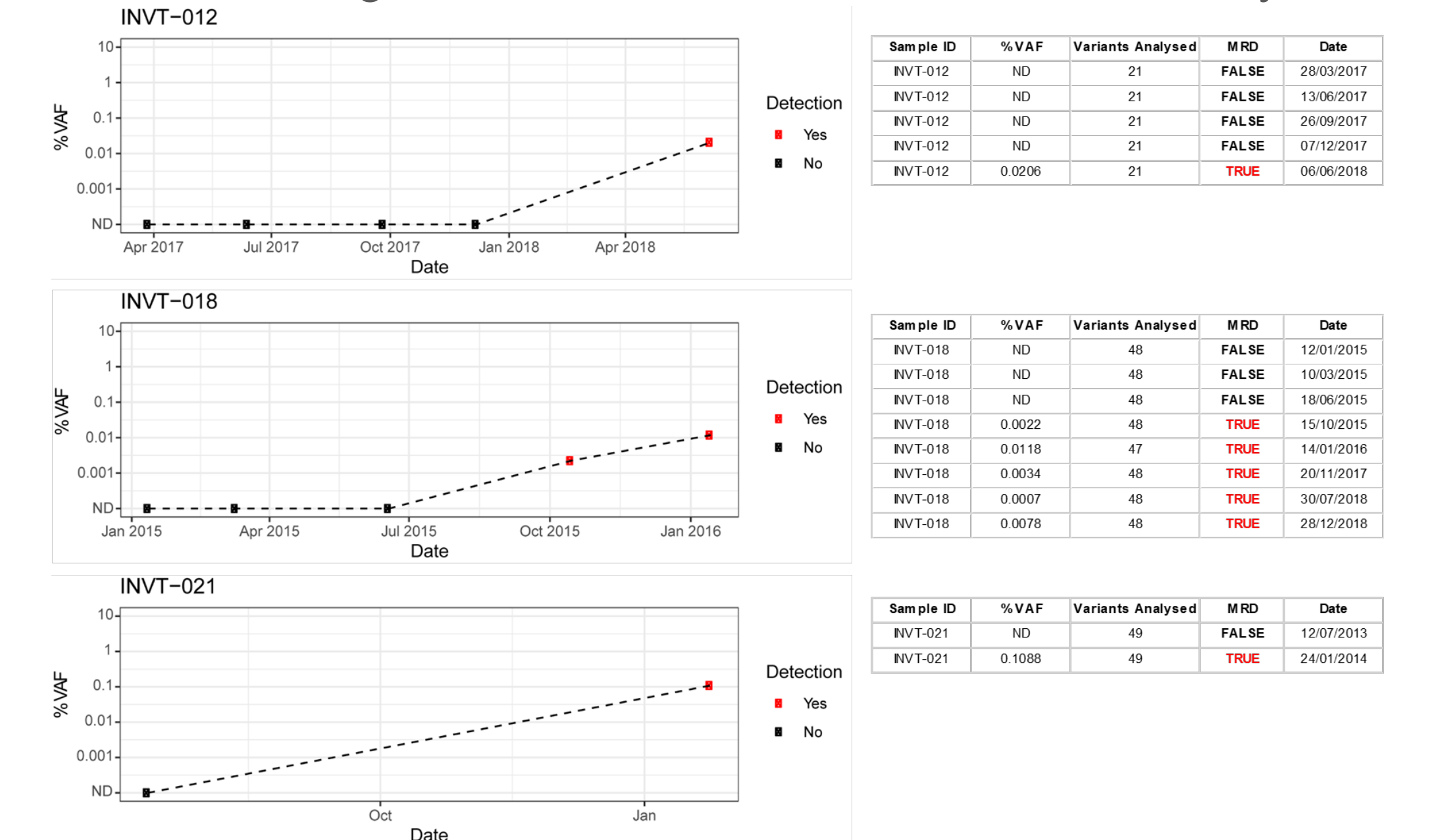
## Tracking clonal evolution in the MRD Setting with ctDNA

In 8/14 patients with multiple tumor samples sequenced, multiple clones (mean 3.4 clones/patient) were identified, with heterogeneous polyclonal relapse in 4/8 patients, and a single clone detectable in 4/8 patients.

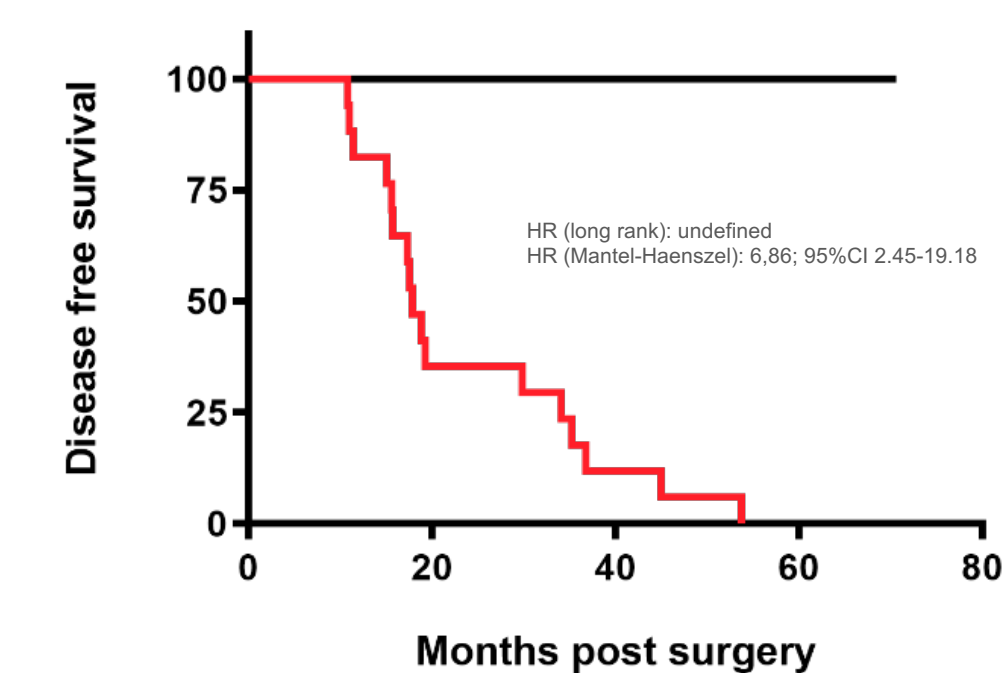


## Brain only relapses detected by RaDaR™ assay

In three patients with brain only relapse, ctDNA was detected prior to relapse in all patients (3/3, 100%) albeit with a reduced lead time over clinical relapse (3.85, 4.21 and 5.65 months), which was not previously achievable with single mutation ddPCR MRD-detection assays.



## Detection of ctDNA Post Surgery is Associated with Higher Risk of Relapse



Median Follow-up: **24.6** months.  
Median Lead Time to clinical Relapse: **8.88 (7.08-9.96)** months.  
Median Lead Time to clinical relapse (extracranial only): **12.89 (11.8-14.02)** months.

## Conclusions

In a proof-of-principle study, ctDNA-detected MRD with personalized sequencing assays associates with relapse free survival and long lead time over clinical relapse in early stage breast cancer. Sequencing based ctDNA testing can detect patients with brain-only relapses and increases sensitivity over first generation ddPCR-based ctDNA assays. Clonal shifts between primary and recurrence can be anticipated at molecular relapse.

## Contact:

Nicholas C Turner ([nicholas.turner@icr.ac.uk](mailto:nicholas.turner@icr.ac.uk))  
Breast Cancer Now Research Centre  
The Institute of Cancer Research, London, UK

## References

- Garcia-Murillas *et al.*, STM (2015)
- Garcia-Murillas *et al.*, JAMA Onc (2019)